

Reference Data

¹³C NMR Spectral Assignments of Some Cyclopentyluracils and 5-Halouracils

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The ¹³C NMR spectra of several 1-(2-hydroxymethylcyclopentyl)- and 1-(2-hydroxymethylcyclopentylmethyl)uracils and 5-halouracils (X = Cl, Br or I) were fully assigned with the aid of one- (¹³C, ¹H-¹H NOE, DEPT) and two-dimensional (HMQC) NMR experiments. © 1997 by John Wiley & Sons, Ltd.

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INTRODUCTION

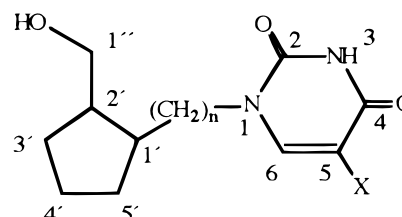
Carbocyclic analogues of nucleosides form an interesting class of biologically active compounds, several of which are potent antitumour and/or antiviral agents.^{1,2} For some years, we have been investigating a group of such analogues, 1,2-disubstituted carbocyclic nucleosides, in which the hydroxymethyl group and heterocyclic base are attached to contiguous carbons of the carbocycle.³ In continuance of our work on the elucidation of the structures of these analogues,⁴ we now report the fully assigned ¹³C NMR spectra of a series of pharmacologically interesting uracils bearing a 2-hydroxymethylcyclopentyl moiety (Fig. 1). This work supplements the ¹³C NMR spectra data available for uracil, uridine⁵ and various uridine analogues.⁶ Additionally, one- and two-dimensional NMR spectroscopic techniques were used to examine the effects on the carbocycle of a halogen (Cl, Br or I) substituent at position 5 of the uracil ring, of a methyl group between the base and the pseudo-glycosidic bond and of a *cis* or *trans* relationship between the base and the 2-hydroxymethyl group.

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RESULTS AND DISCUSSION

Table 1 lists the ¹³C chemical shift data for compounds 1–12.

The signals due to the cyclopentyl ring carbons were fully assigned by comparison of the ¹³C NMR spectra of compounds 1–12 with the corresponding heteronuclear multiple-quantum correlation (HMQC) spectra of 2-hydroxymethylcyclopentyls bearing a guanine or 2,6-diaminopurine base.⁷ As would be expected, C-1' was heavily



Compound	n	Isomer	X
1	0	<i>cis</i>	H
2	0	<i>trans</i>	H
3	1	<i>cis</i>	H
4	1	<i>trans</i>	H
5	0	<i>cis</i>	Cl
6	0	<i>cis</i>	Br
7	0	<i>cis</i>	I
8	0	<i>trans</i>	I
9	1	<i>cis</i>	Cl
10	1	<i>cis</i>	Br
11	1	<i>cis</i>	I
12	1	<i>trans</i>	I

Figure 1. Compounds 1–12.

Table 1. ¹³C NMR chemical shifts (δ, ppm) of compounds 1–12

Compounds	C-1'	C-2'	C-3'	C-4'	C-5'	C-1''	(CH ₂) _n -N	C-2	C-4	C-5	C-6
1	58.1	42.6	27.5	22.5	29.2	61.0	—	151.9	163.6	100.4	143.8
2	59.0	45.8	27.5	22.5	30.9	62.8	—	151.3	163.4	101.4	143.3
3	40.8	43.3	27.6	22.3	28.2	61.1	47.9	151.1	163.7	100.7	145.9
4	41.6	45.2	29.1	24.2	30.2	64.6	51.8	151.5	164.0	100.9	146.3
5	59.2	41.7	27.2	22.5	28.8	61.0	—	151.0	159.5	105.8	141.1
6	59.2	41.7	27.3	22.5	28.9	61.1	—	151.2	159.7	94.3	143.5
7	59.1	41.9	27.3	22.5	29.0	61.1	—	151.6	161.0	67.7	148.0
8	60.1	45.2	27.1	22.1	30.6	62.8	—	150.8	160.6	68.7	147.6
9	41.1	43.7	28.0	22.7	28.5	61.4	48.7	151.0	160.2	106.4	143.2
10	41.1	43.7	28.0	22.7	28.5	61.4	48.7	150.9	159.9	94.8	145.8
11	40.8	43.4	27.6	22.3	28.1	61.1	48.2	150.9	161.0	67.8	150.1
12	43.4	45.3	29.2	24.8	31.5	66.6	53.9	151.5	161.0	67.9	149.8

Reference Data

deshielded by the heterocycle: in the 2-hydroxycyclopentyl compounds (1, 2 and 5–8) the C-1' signal appeared at 59.1 ± 1 ppm and in the 2-hydroxycyclopentylmethyl compounds (3, 4 and 9–12) it was shifted to higher frequencies, appearing at 42.1 ± 1.3 ppm, so that C-2' (43.5 ± 1.8 ppm) became the most deshielded cyclopentyl ring carbon. The remaining cyclopentyl ring carbons were only slightly affected by changes in the stereochemistry and/or substituent at position 5 of the uracil ring: the C-5' signal appeared at 29.8 ± 1.7 ppm, very close to the C-3' signal at 28.1 ± 1 ppm, and the C-4' signal appeared at 23.5 ± 1.4 ppm.

The signals due to the uracil ring carbons were mainly assigned by comparison of their chemical shifts with these data for uridine;⁵ nonetheless, to distinguish between the closely lying signals due to the C-2 quaternary and the C-2 methine groups of the 5-halouracils (5–12), DEPT experiments were also carried out. The effects of the halogen substituents are evident from the ^{13}C chemical shift data: deshielding at C-5 increases in the order $\text{I} < \text{Br} < \text{Cl}$, whereas at C-6 it increases in the order $\text{Cl} < \text{Br} < \text{I}$. Deshielding by the 5-halo substituent was also evident in the ^1H NMR spectra, in which the NH (3) and C-6 signals of compounds 5–12 had chemical shifts approximately 0.5 ppm higher than in the corresponding uracil compounds (1–4). For the latter uracil compounds and the 5-ioduracil compounds (7, 8, 11 and 12), there were only minor differences between the chemical shifts for the uracil ring carbons of *cis* and *trans* isomers.

EXPERIMENTAL

Compounds 1–4 were synthesized by constructing the uracil base about an appropriate preformed amino alcohol.⁷ The amino alcohol was first reacted with 3-ethoxy-2-propenyl isocyanate in DMF at -20°C and then the resulting urea was cyclized in sulphuric acid^{8,9} (compounds 1 and 2 were separated by flash chromatography on silica gel, with CHCl_3 as eluent). The 5-halouracil compounds (5–12) were obtained by treatment of compounds 1–4 with iodine and HNO_3 in dioxane,¹⁰ or with *N*-chloro- or *N*-bromosuccinimide in acetic acid.¹¹ All compounds were fully characterized, both physically and spectroscopically, and their stereochemistry was determined by means of ^1H – ^1H nuclear Overhauser effect (NOE) experiments by irradiation of CH (1').^{3,4,7}

^{13}C and ^1H NMR spectra of samples as approximately 10% solutions in $\text{DMSO}-d_6$ were recorded at room temperature in 5 mm o.d. tubes. The chemical shifts were internally referenced to TMS (0 ppm).

One-dimensional ^{13}C NMR spectra were recorded on a Bruker AMX 300 NMR spectrometer operating at 75.47 MHz, typically with a 30° pulse flip angle, a pulse repetition time of 1.8 s and a spectral width of 17857 Hz with 32K data points. For the DEPT sequence, the width of the 90° pulse for ^{13}C was 4 μs and that of the 90° pulse for ^1H was 9.5 μs ; the delay $2J_{\text{C-H}}^{-1}$ was set at 3.45 ms.

^1H NMR and homonuclear NOE¹² experiments were performed on a Bruker WM-250 Fourier transform spectrometer operating at 250.13 MHz, typically with a 30° pulse flip angle, a pulse repetition time of 2 s and a spectral width of 2726 Hz with 16K data points.

^1H -detected, one-bond HMQC spectra were recorded on a Bruker AMX 500 spectrometer using a pulse sequence (the INV4GS micro program of the Bruker software) that allowed gradient selection. Spectra were collected in the t_1 domain in 256 experiments with 2K data points, and spectral widths of 5050 and 27669 Hz in the F_2 (^1H) and F_1 (^{13}C) dimensions, respectively. The relaxation delay, D_1 , was set to 2 s and D_2 was empirically optimized to 3.5 ms. Data were processed using sine-bell weighting functions in both dimensions.

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